



Antimicrobial effects of *Nigella sativa* L.: Review

S. Derbal, *, A Djekouna^{1,2}

^{1,2} Pharmaceutical Sciences Research Center (CRSP),
Constantine 25000, Algeria
saidderbal19@gmail.com

Abstract. In recent decades, the phenomenon of resistance of microorganisms to anti-infectious molecules used in therapy has become a health problem throughout the world. Therefore, it is necessary to adopt new approaches to overcome this phenomenon of resistance with the suggestion of finding the solution by researching bioactive molecules in medicinal plants and exploiting them in the pharmaceutical industry in order to develop new antimicrobial drugs. *Nigella sativa* is used in many regions in traditional medicine, has broad therapeutic effects, and has been reported to have significant antimicrobial effects. This review article attempts to describe some antimicrobial effects such as antiviral, antibacterial, antifungal and antiprotozoal effects, and which have been carried out by various researchers.

Keywords: *Nigella sativa*, Antiviral, Antibacterial, Antifungal, Antiprotozoal

1 Introduction

Nigella sativa, known as black seed or black cumin in English and habbat al-barakah in Arabic. *N. sativa* is an annual herbaceous plant in family Ranunculaceae which grows well on loamy soils in tropical and subtropical regions. Also, in Asia, Europe, the Middle East and other Mediterranean countries, *N. sativa* has been regularly and abundantly cultivated for various purposes [1].

N. sativa is widely used natural remedy and the seeds are extensively used as spice, carminative, condiment and aromatic. Traditionally, they have been used as diuretic, diaphoretic, stomachic, liver tonic, digestive, emmenagogue and galactagogue. Also, they are used in diarrhoea, indigestion, dyspepsia, sour belching, loss of appetite, vomiting, dropsy, puerperal diseases [2].

The seeds of *N. sativa* are composed of 66.5% linoleic acid (18: 2) and 23.5% oleic acid (18: 1) as the main fatty acids. Their content of trace elements like Co, Ni, Fe, Zn, Cu, Mn and Cr is 0.12, 1.48, 117.32, 41.42, 30.26, 28.56 and 2.55 µg/g respectively, and their vitamin content is 10.19 µg/g for -tocopherol, 2.28 µg/g for -tocopherol, 0.18 µg/g for retinol, 1.38 µg/g for vitamin D2, 1.85 µg/g for vitamin K1 and 2.15 µg/g for vitamin K2 [3]. *N. sativa* is rich in

* Corresponding author

Received August 15, 2021; accepted September 18, 2021.

steroids, alkaloids, flavonoids, glycosides, saponins and terpenoids [4]. The main compounds of the fixed oil and essential oil of *N. sativa* are thymoquinone, p-cymene and longifolene [5, 6]. Also, oleoresins extracted in different solvents (ethanol, ethyl acetate and n-hexane) contain linoleic acid as a major component [7].

Various pharmacological aspects of *N. sativa* extracts have been reported, including its anti-inflammatory [8, 9], analgesic and antipyretic [8], Antipsoriatic [10], Antitumor [11], immunopotentiating [12], Antioxidant [13], Antidiabetic [14], Hepatoprotective [15], Neuroprotective [16], Anti-ischemic [17], Gastroprotective [18], antihypertensive [19], Diuretic and hypotensive [20] properties.

This review investigates the antimicrobial effects of *N. sativa* of which the most important antiviral, antibacterial, antifungal and antiprotozoal effects are discussed below. It is not possible to discuss all of its antimicrobial effects here because the literature on this plant is very extensive.

2 Antiviral activity

In a study of patients with hepatitis C ineligible for interferon (IFN)-, administration of *N. sativa* was tolerable, safe, decreased HCV viral load and improved oxidative stress, clinical and glycemic control in diabetic patients [21]. In another study, administration of ethanolic extract of *N. sativa* to patients with HCV showed potential therapeutic benefits via the decrease in viral load and the relief of impaired liver function, with an effect more powerful offered by their mixture with the ethanolic extract of *Zingiber officinale* [22].

Dithymoquinone (DTQ) the major constituent of *N. sativa* showed in a study a strong potential for binding to the SARS-CoV-2: ACE2 interface and could therefore be predicted as a plausible inhibitor to disrupt viral-host interactions [23]. Also, it was reported in a study that an adult person diagnosed with HIV positive became HIV negative after receiving treatment with *N. sativa* for several months, since this case is considered surprising [24].

N. sativa is of importance in agriculture since in in-vitro experience, decoction and infusion of *N. sativa* seeds inhibited the symptoms of Zucchini Yellow Mosaic Virus (ZYMV) on squash plants by 85% and 80%, respectively. In post-experiment, decoction and infusion gave 70% and 65% inhibition of ZYMV, respectively. In addition, the soaking of pumpkin seeds in decoction showed higher activity against ZYMV than the infusion. The decoction showed a maximum percentage of viral inhibition (95%) after soaking for 24 hours, while the infusion gave 90% inhibition after soaking for 24 hours [25]. In another study, the volatile oil and acetone extract of *N. sativa* showed effective inhibitory activity against papaya ringspot virus (PRSV), using *Chenopodium amaranticolor* coste and Reyn, a host of the lesion [26].

Moreover, *N. sativa* can be exploited in the veterinary field since in an experiment, the ethanolic extract of black seed was used in the embryonated eggs inoculated with the Newcastle disease virus (NDV), it increased the level of survival, and gross and histopathological lesions were not intense in embryos treated with

this extract, which justifies its antiviral and immunotherapeutic effect against NDV infection [27]. Also, *N. sativa* has shown antiviral activity against infectious laryngotracheitis virus (ILTV) [28]. In another experiment, supplementation of the commercial diet with seeds of *N. sativa* resulted in turkeys experimentally infected with the H9N2 avian influenza virus a considerable improvement in immune reactivity and suppression of the pathogenicity of this virus [29]. Also, intraperitoneal administration of *N. sativa* oil to BALB/c mice experimentally infected with murine cytomegalovirus (MCMV) showed an effective antiviral effect which may be mediated by increased number and function of Mf and IFN-production [30]. In addition, treatment with alcoholic extract of *N. sativa* of Vero cell lines infected with PPRV resulted in an increase in the viability of the infected cells and a reduction in the cytopathic effects of PPRV [31].

3 Antibacterial activity

N. sativa essential oil and its compounds (thymoquinone and carvacrol) have shown substantial antibacterial activity against *Listeria monocytogenes*, and the essential oil may even have potential to control antibiotic resistance in *Listeria* [32], and even *N. sativa* oil has shown strong antibacterial activity against twenty strains of *Listeria monocytogenes* [33]. Also, *N. sativa* oil was found to be more effective against several species of *Salmonella* for which even ceftriaxone and ciprofloxacin were ineffective [6]. In another study, the essential oil and methanolic extract of *N. sativa* had antibacterial activity in-vitro against *Salmonella enterica* isolates of human and poultry origin resistant to antibiotics, in contrast the aqueous extract had no anti-salmonella activity [34].

In a study, aqueous and ethanolic extracts of *N. sativa* inhibited the growth of methicillin-resistant *Staphylococcus aureus* (MRSA) [4] and in another study an extract of *N. sativa* prepared by the method reflux extraction had an inhibitory effect on strains of MRSA [35].

N. sativa essential oil and its compound thymoquinone have demonstrated significant anticariogenic activity against 30 clinical cariogenic bacteria. The strongest activity of the essential oil was observed against *Streptococcus mitis*, *Streptococcus mutans*, *Streptococcus constellatus* and *Gemella haemolysans*. However, thymoquinone was active against all the strains studied, in particular *Streptococcus mutans* and *Streptococcus mitis* [36]. The seed methanolic extract of *N. Sativa* was more effective than the ether extract in inhibiting the growth of two cariogenic bacteria *Streptococcus mutans* and *Streptococcus mitis* [37].

N. sativa is of great importance in dermatology since more than half of the 19 multidrug-resistant *Staphylococcus aureus* isolates isolated from diabetic wounds were sensitive to different concentrations of *N. sativa* oil [38]. In a study designed to explore the antimicrobial effect of *N. sativa* seed extract against skin pustules, the antibacterial effect of *N. sativa* seed extract on skin pustules with staphylococcal infections was evaluated in-vivo and compared to mupirocin in newborns, this extract was as effective as mupirocin and without any side effects [39]. In another study, three crude extracts (methanolic, hexane and ethyl acetate) of *N.*

sativa seeds were evaluated for their anti-acne potential against certain bacterial species responsible for acne; among these three crude extracts tested, the highest antibacterial activity against *Staphylococcus aureus* was observed in the ethyl acetate extract. Subsequently, topical gels were formulated incorporating the ethyl acetate extract at three different concentrations. These three formulations were able to inhibit the growth of *Staphylococcus aureus* and *Propionibacterium acnes*, with the highest antibacterial activity in the formulation comprising 15% of the seed extract [40].

N. sativa seed oil has been shown to have bactericidal activity in-vitro against bacteria commonly involved in otitis media and otitis externa (clinical isolates of *Streptococcus pneumoniae*, *Moraxella catarrhalis*, and *Haemophilus influenzae*). But this oil has not been shown to be effective against clinical isolates of *Pseudomonas aeruginosa* [41].

The seed crude extract of *N. sativa* has demonstrated antibacterial activity against *Bacillus subtilis*, *Klebsiella pneumoniae*, *Mycobacterium phlei* and methicillin-resistant and methicillin-sensitive *Staphylococcus aureus* [42]. Also, 37 isolates of *Shigella dysenteriae* 1, *Shigella flexneri*, *Shigella sonnei* and *Shigella boydii* and 10 strains of *Vibrio cholerae* and *Escherichia coli* showed promising sensitivity to volatile oil of *N. sativa* [43]. In another study, the fixed and volatile oils of *N. Sativa* showed antibacterial activity in-vitro against all standard strains (ATCC) of *Escherichia coli*, *Pseudomonas aeruginosa*, *Acinetobacter baumannii*, *Staphylococcus aureus* and *Enterococcus faecalis*, and anti-mycobacterial activity against two standard strains of *Mycobacterium tuberculosis* H37Rv and *Mycobacterium avium* (ATCC). Usually, volatile oil was much more efficient than fixed oil. On the other hand, these oils were judged ineffective against the isolated bacterial strains [44].

Thymoquinone (TQ) the major active ingredient of *N. sativa* seed has shown significant antimicrobial activity against standard strains (ATCC) of four anaerobic bacteria which are *Clostridium difficile*, *Clostridium perfringens*, *Bacteroides fragilis* and *Bacteroides thetaiotaomicron* [45]. In another study, thymoquinone showed antibacterial activity against eleven reference bacterial strains considered to be human pathogens and which are Gram-negative bacilli: *Escherichia coli* ATCC 35218, *Salmonella enterica* serovar Typhimurium ATCC 14028, *Pseudomonas aeruginosa* ATCC 27853, *Vibrio alginolyticus* ATCC 33787, *Vibrio paraheamolyticus* ATCC 17802; Gram-positive bacilli: *Bacillus cereus* ATCC 14579, *Listeria monocytogenes* ATCC 19115 and Gram-positive cocci: *Enterococcus faecalis* ATCC 29212, *Micrococcus luteus* NCIMB 8166, *Staphylococcus aureus* ATCC 25923, *Staphylococcus epidermidis* CIP 106510. In addition, it has shown that it has antibiofilm potential against *Pseudomonas aeruginosa* ATCC 27853, *Enterococcus faecalis* ATCC 29212, *Staphylococcus aureus* ATCC 25923 and *Staphylococcus epidermidis* CIP 106510 [46].

In a study aimed at evaluating the antibacterial activity of total crude extracts and essential oil of *N. sativa* seed in male mice infected intraperitoneally with *Staphylococcus aureus* or *Escherichia coli*; the methanolic and chloroform extracts as well as the essential oil had dose-dependent antibacterial activities

on these bacteria, while the aqueous extract showed no inhibitory effects on any of these bacteria [47].

4 Antifungal activity

The essential oil and various extracts (aqueous and methanolic) of *N. sativa* and especially thymoquinone have demonstrated powerful antifungal effects on pathogenic dermatophyte strains which are *Trichophyton mentagrophytes*, *Microsporum canis* and *Microsporum gypseum* [5]. Also, the seed ether extract of

N. sativa and its active ingredient thymoquinone demonstrated antifungal activity against eight species of dermatophytes: four species of *Trichophyton rubrum* and one of *Trichophyton interdigitale*, *Trichophyton mentagrophytes*, *Epidermophyton floccosum* and *Microsporum canis* [48]. In another study, the essential oil of *N. sativa* demonstrated strong anti-dermatophytic activity against *Microsporum gypseum*, *Trichophyton rubrum*, *Trichophyton simii*, *Chrysosporium tropicum* and *Chrysosporium evolceanui* [49].

N. sativa can be used to treat candidiasis since *N. sativa* seed oil had strong antifungal activity against several isolates of *Candida albicans*, *Candida dubliniensis*, *Candida glabrata* and *Candida krusei* [50] and even ethanolic extract of *N. sativa* has demonstrated effective antifungal activity against *Candida albicans* [51]. In an in-vivo study, the seed aqueous extract of *N. sativa* exhibited an inhibitory effect against experimental candidiasis caused by *Candida albicans* in mice, in which it considerably inhibited the growth of this pathogen in the kidneys, liver and spleen [52].

Different fractions (aqueous, chloroform, n-hexane, ethyl acetate and n-butanol) of the methanolic extract of *N. sativa* vegetative parts inhibited the production of fungal biomass from two soil fungal species, *Fusarium oxysporum* and *Macrophomina phaseolina* [53]. Also, the oil and oleoresins (ethanolic, ethyl acetate and n-hexane) of *N. sativa* have demonstrated inhibitory activity against *Aspergillus niger*, *Aspergillus flavus*, *Fusarium moniliforme*, *Fusarium graminearum* and *Penicillium viridicatum* [7]. In addition, the essential oil of *N. sativa* has shown fungicidal activity against eight fungi which are *Aspergillus niger*, *Aspergillus flavus*, *Fusarium oxysporum*, *Fusarium moniliforme*, *Fusarium nivale*, *Fusarium semitectum*, *Drechslera hawaiiensis* and *Alternaria alternata* [54].

5 Antiprotozoal activity

In an in-vitro study aimed to evaluate the anti-leishmanial effects of *N. sativa* against *Leishmania tropica* and *Leishmania infantum*; the essential oil, the methanolic extract and in particular thymoquinone, had a powerful antileishmanial activity on the promastigotes of these two species since these extracts induced a significant decrease in the growth rate of the amastigote forms of the two species [55]. In another in-vitro study, the essential oil of

N. sativa and its main bioactive component thymoquinone showed cytotoxic effects on promastigotes of *Leishmania tropica* [56] and even the chloroform extract of *N. sativa* seed showed effective leishmanicidal activity against this *Leishmania* species in an in-vitro model [57]. In addition, the aqueous extract of the *N. sativa* plant has shown a leishmanicidal effect in-vitro against *Leishmania major* [58].

N. sativa is effective in controlling intestinal-tropic protozoa since in an experiment, supplementation of the diet with 1% whole crushed seeds of *N. sativa* resulted in broilers experimentally infected with the field strain *Eimeria tenella* improved biochemical parameters and cecal lesions and reduced oocyst excretion [59]. Also, the oil and alcoholic extract of *N. sativa* seed resulted in reduced oocyst excretion of *Cryptosporidium* spp. in Swiss Albino mice experimentally infected with these protozoa [60]. In other in-vitro studies, the ethanolic extract of *N. sativa* seed was shown to have potent antiprotozoal activity against the trophozoites of *Entamoeba histolytica* and *Giardia lamblia* [61, 62]. In addition, thymoquinone the active component of *N. sativa* has been shown to have more potent in-vitro anti-protozoan activity against *Entamoeba histolytica* than against *Giardia lamblia* [63].

N. sativa oil showed a trypanocidal effect in rats experimentally infected with *Trypanosoma brucei* as there was a reduction in parasitaemia and an extension of the lifespan of these rats [64]. Also, aqueous and ethanolic extracts of *N. sativa* seed showed antitrypanosomal activity in an in-vitro test against *Trypanosoma evansi* [65].

In albino mice experimentally infected with the virulent *Toxoplasma RH* strain, *N. sativa* oil alone has significant immunostimulatory and antioxidant properties and it has no direct anti-toxoplasma effect, whereas its combination with pyrimethamine (PYR) produced a potent therapeutic effect since it considerably increased the survival rate and decreased parasite density and pathological damage in the liver and spleen [66]. In another experiment, *N. sativa* oil showed promising prophylactic and therapeutic effects on murine toxoplasmosis in mice experimentally infected with brain cysts of *Toxoplasma gondii* (strain Me49). It dramatically improved the protection of infected mice from death and reduced the burden of brain cysts. The brains of mice that received *N. sativa* oil showed milder lesions and higher iNOS expression [67].

In Swiss albino mice experimentally infected with *Plasmodium yoelli nigeriensis* (*Plasmodium yoelli*), the methanolic extract of *N. sativa* seed was found to be more effective than chloroquine (CQ) in clearing the parasite and restoring biochemical indices modified by infection [68]. The methanolic and ethyl acetate extracts of *N. sativa* showed good in-vitro activity against the Chloroquine Sensitive Sierra Leone 1 (D6) and Chloroquine-resistant Indochina 1 (W2) strains of *Plasmodium falciparum*, and in-vivo against the ANKA strain of *Plasmodium berghei* [69]. Also, intraperitoneal and oral administrations of ethanolic, aqueous and chloroform extracts of *N. sativa* seed in mice experimentally infected with *Plasmodium berghei* showed suppression activity since they decreased parasitaemias and increased survival times of infected mice [70]. In another ex-

periment, the methanolic and ethyl acetate extracts of *N. sativa* seed showed in-vivo antimalarial activity against the *Plasmodium berghei* ANKA strain by the parasitaemia suppression test in a mouse model [71]. Also, seeds and oil of *N. sativa* have shown in-vivo antimalarial activity against chloroquine-sensitive *Plasmodium berghei* NK65 strain by parasitaemia suppression assay in albino mice, and they have potential to improve the efficacy of chloroquine (CQ) [72]. In addition, the aqueous extract of *N. sativa* seed increased the survival rate and induced a reduction in the number of parasitaemias and the level of nitric oxide in mice infected with the parasite *Plasmodium berghei* NK65 [73].

The alcoholic extract, the aqueous extract and the seed oil of *N. sativa* showed in-vitro inhibitory activity on the growth and activity of *Trichomonas vaginalis* [74, 75, 76]. Also, *N. sativa* oil showed a high toxic effect in-vitro on trophozoites of *Trichomonas vaginalis* since it induced severe cell damage with cytoplasmic and nuclear destruction, while the effect of alcoholic extract of *N. sativa* was moderate [77].

Finally, the aqueous extract of *N. sativa* has been shown to be effective in the treatment of experimentally induced *Acanthamoeba keratitis* (AK) [78].

The difference in antimicrobial activity between the different extracts can be explained by several factors since the physicochemical properties of *N. sativa* extracts vary according to regions or geographic areas [79] and the method of extraction [80]. Also, the anti-microbial potential of *N. sativa* seed extracts varies according to regions or geographic areas [81, 82]. In addition, the chemical composition and antimicrobial activity of *N. sativa* seed essential oils and thymoquinone content vary depending on the extraction methods [83, 84].

The whole plant, seeds, extracts and bioactive molecules of *Nigella sativa* were concerned for the study of the antimicrobial effects of which these studies were carried out by different methods: in-vitro and in-vivo (patients, farm animals and experimental animals). Figure 1 represents a summary of the studies mentioned in this review.

6 Conclusion

The black seed is a well-documented medicinal plant and the present study showed that *N. sativa* extracts have a broad spectrum of antimicrobial activity. It is a pity that this miracle medicinal plant is not more widely exploited in public health, veterinary and agricultural field since it can be used as a readily available source of antimicrobial molecules in food, pharmaceutical and cosmetic products.

As many experiments illustrate its potential for many infectious diseases, more clinical and pathological studies must be carried out to investigate the untapped potential of this plant. Thus, more work is needed to determine the bioactive molecules, the mechanisms of action, the dosage and the ideal doses for the different extracts.

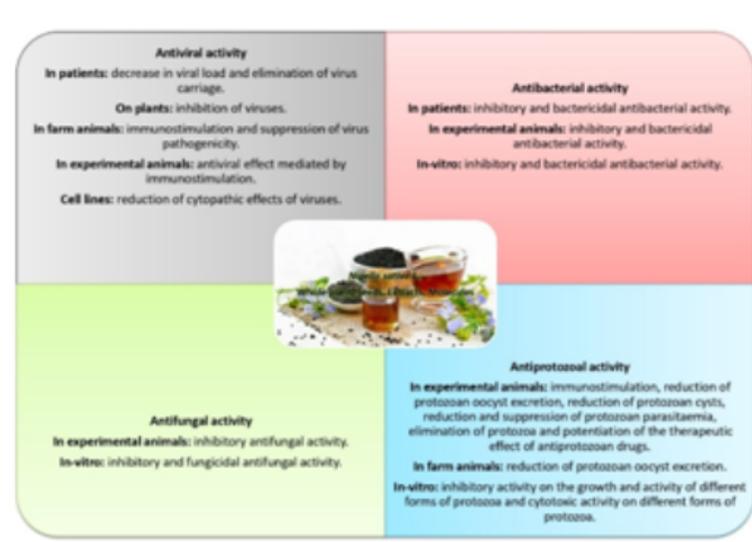


Fig. 1. Summary diagram of the antimicrobial effects of *N. sativa*.

References

1. Majeed A., Muhammad Z., Ahmad H., Rehmanullah, Hayat S.S.S., Inayat N., Siyyar S., *Nigella sativa* L.: Uses in traditional and contemporary medicines – An overview. *Acta Ecol. Sin.*, (2020), 1-6.
2. Gilani A.H., Jabeen Q., Khan M.A.U., A review of medicinal uses and pharmacological activities of *Nigella sativa*. *Pak. J. Biol. Sci.*, (2004), 7(4): 441-451.
3. [Vatansev H., Ciftci H., Ozkaya A., Ozturk B., Evliyaoglu N., Kiyici A., Chemical composition of *Nigella sativa* L. seeds used as a medical aromatic plant from east Anatolia region, Turkey. *Asian J. Chem.*, (2013), 25(10): 5490-5492.
4. Zakka A.W., Ashefo D.P., Abraham O.J., Wurtu J.R., Yaki L.M., Muhammad R., Antibacterial activity of crude ethanolic extract of *Nigella sativa* seed on molecularly identified methicillin resistant clinical isolate of *Staphylococcus aureus*, in Kaduna State, Nigeria. *AJMAB*, (2019), 4(1): 9-15.
5. Mahmoudvand H., Sepahvand A., Jahanbakhsh S., Ezatpour B., Ayatollahi Mousavi S.A., Evaluation of antifungal activities of the essential oil and various extracts of *Nigella sativa* and its main component, thymoquinone against pathogenic dermatophyte strains. *J. Mycol. Med.*, (2014a), 24: 155-161.
6. Sarwar A., Latif Z., GC-MS characterisation and antibacterial activity evaluation of *Nigella sativa* oil against diverse strains of *Salmonella*. *Nat. Prod. Res.*, (2015), 29(5): 447-451.
7. Singh S., Das S.S., Singh G., Schuff C., de Lampasona M.P., Catalán C.A.N., Composition, in vitro antioxidant and antimicrobial activities of essential oil and oleoresins obtained from black cumin seeds (*Nigella sativa* L.). *Biomed. Res. Int.*, (2014), 1-10.
8. Al-Ghamdi M.S., The anti-inflammatory, analgesic and antipyretic activity of *Nigella sativa*. *J. ethnopharmacol.*, (2001), 76: 45-48.

9. Chehl N., Chipitsyna G., Gong Q., Yeo C.J., Arafat H.A., Anti-inflammatory effects of the *Nigella sativa* seed extract, thymoquinone, in pancreatic cancer cells. *HPB*, (2009), 11: 373-381.
10. Dwarampudi L.P., Palaniswamy D., Nithyanantham M., Raghu P.S., Antipsoriatic activity and cytotoxicity of ethanolic extract of *Nigella sativa* seeds. *Pharmacogn. Mag.*, (2012), 8(32): 268-272.
11. Musa D., Dilsiz N., Gumushan H., Ulakoglu G., Bitiren M., Antitumor activity of an ethanol extract of *Nigella sativa* seeds. *Biologia Bratislava*, (2004), 59(6): 735-740.
12. Swamy S.M.K., Tan B.K.H., Cytotoxic and immunopotentiating effects of ethanolic extract of *Nigella sativa* L. seeds. *J. Ethnopharmacol.*, (2000), 70: 1-7.
13. Meziti A., Meziti H., Boudiaf K., Mustapha B., Bouriche H., Polyphenolic profile and antioxidant activities of *Nigella Sativa* seed extracts in vitro and in vivo. *World Academy of Science, Engineering and Technology. Int. J. Biotechnol. Bioeng.*, (2012), 6(4): 109-117.
14. Benhaddou-Andaloussi A., Martineau L.C., Spoor D., Vuong T., Leduc C., Joly E., Burt A., Meddah B., Settaf A., Arnason J.T., Prentki M., Haddad P.S., Antidiabetic Activity of *Nigella sativa* seed extract in cultured pancreatic -cells, skeletal muscle cells, and adipocytes, *Pharmaceutical Biology.*, (2008), 46(1-2): 96-104.
15. Adam G.O., Rahman M.M., Lee S.J., Kim G.B., Kang H.S., Kim J.S., Kim S.J., Hepatoprotective effects of *Nigella sativa* seed extract against acetaminophen-induced oxidative stress. *Asian Pac. J. Trop. Med.*, (2016), 9(3): 221-227.
16. Islam M.H., Ahmad I.Z., Salman M.T., Neuroprotective effects of *Nigella sativa* extracts during germination on central nervous system. *Pharmacogn. Mag.*, (2015), 11(42): 182-189.
17. Hosseinzadeh H., Jaafari M.R., Khoei A.R., Rahmani M., Anti-ischemic effect of *Nigella sativa* L. seed in male rats. *IJPR*, (2006), 1: 53-58.
18. Rifat-uz-Zaman, Akhtar M.S., Khan M.S., Gastroprotective and anti-secretory effect of *Nigella sativa* seed and its extracts in indomethacin-treated Rats. *Pak. J. Biol. Sci.*, (2004), 7 (6): 995-1000.
19. Jaarin K., Foong W.D., Yeoh M.H., Kamarul Z.Y., Qodriyah H.M., Azman A., Zuhair J.S.F., Juliana A.H., Kamisah Y., Mechanisms of the antihypertensive effects of *Nigella sativa* oil in L-NAME-induced hypertensive rats. *Clinics*, (2015), 70(11): 751-757.
20. Zaoui A., Cherrah Y., Lacaille-Dubois M.A., Settaf A., Amarouch H., Hassar M., Diuretic and hypotensive effects of *Nigella sativa* on the spontaneously hypertensive rat. *Thérapie*, (2000), 55(3): 379-382.
21. Barakat E.M.F., El Wakeel L.M., Hagag R.S., Effects of *Nigella sativa* on outcome of hepatitis C in Egypt. *World J. Gastroenterol.*, (2013), 19(16): 2529-2536.
22. Abdel-Moneim A., Morsy B.M., Mahmoud A.M., Abo-Seif M.A., Zanyaty M.I., Beneficial therapeutic effects of *Nigella sativa* and/or *Zingiber officinale* in HCV patients in egypt. *EXCLI Journal*, (2013), 12: 943-955.
23. Ahmad S., Abbasi H.W., Shahid S., Gul S., Abbasi S.W., Molecular docking, simulation and MM-PBSA studies of *Nigella sativa* compounds: a computational quest to identify potential natural antiviral for COVID-19 treatment. *J. Biomol. Struct. Dyn.*, (2020), 1-9.
24. Onifade A.A., Jewell A.P., Adedeji W.A., *Nigella sativa* concoction induced sustained seroreversion in HIV patient. *Afr. J. Tradit. Complement. Altern. Med.*, (2013), 10(5): 332-335.
25. El-Shafi S.A., Preliminary Studies on Antibacterial and Antiviral Activities of Five Medicinal Plants. *J. Plant. Pathol. Microb.*, (2013), 4: 190.

26. Maurya S., Marimuthu P., Singh A., Rao G.P., Singh G., Antiviral activity of essential oils and acetone extracts of medicinal plants against Papaya Ring Spot Virus. *J. Essent. Oil Bear. Pl.*, (2005), 8(3): 233-238.
27. Khan A.U., Tipu M.Y., Shafee M., Khan N.U., Kiani M.M.T., Rafeeq M., Shah S.I.A., In-ovo antiviral effect of *Nigella sativa* extract against Newcastle Disease Virus in experimentally infected chicken embryonated eggs. *Pak. Vet. J.*, (2018).
28. Zaher K.S., Ahmed W.M., Zerizer S.N., Observations on the biological effects of black cumin seed (*Nigella sativa*) and green tea (*Camellia sinensis*). *Glob. Vet.*, (2008), 2(4): 198-204.
29. Umar S., Munir M.T., Subhan S., Azam T., Nisa Q., Khan M.I., Umar W., Rehman Z., Saqib A.S., Shah M.A., Protective and antiviral activities of *Nigella sativa* against avian influenza (H9N2) in turkeys. *J. Saudi Soc. Agric. Sci.*, (2016).
30. Salem M.L., Hossain M.S., Protective effect of black seed oil from *Nigella sativa* against murine cytomegalovirus infection. *Int. J. Immunopharmacol.*, (2000), 22: 729-740.
31. Aqil K., Khan M.R., Aslam A., Javeed A., Qayyum R., Yousaf F., Yasmeen F., Sohail M.L., Umar S., In vitro Antiviral activity of *Nigella sativa* against Peste des Petits Ruminants (PPR) Virus. *Pakistan J. Zool.*, (2018), 50(6): 2223-2228.
32. Mouwakeh A., Telbisz A., Spengler G., Mohácsi-Farkas C., Kiskó G., Antibacterial and resistance modifying activities of *Nigella sativa* essential oil and its active compounds against *Listeria monocytogenes*. *in vivo*, (2018), 32: 737-743.
33. Nair M.K.M., Vasudevan P., Venkitanarayanan K., Antibacterial effect of black seed oil on *Listeria monocytogenes*. *Food Control.*, (2005), 16: 395-398.
34. Ashraf S., Anjum A.A., Ahmad A., Firyal S., Sana S., Latif A.A., In vitro activity of *Nigella sativa* against antibiotic resistant *Salmonella enterica*. *Environ. Toxicol. Pharmacol.*, (2018), 58: 54-58.
35. Hannan A., Saleem S., Chaudhary S., Barkaat M., Arshad M.U. Antibacterial activity of *Nigella sativa* against clinical isolates of methicillin resistant staphylococcus aureus. *J. Ayub. Med. Coll. Abbottabad.*, (2008), 20(3): 72-74.
36. Jrah Harzallah H., Kouidhi B., Flamini G., Bakhrouf A., Mahjoub T., Chemical composition, antimicrobial potential against cariogenic bacteria and cytotoxic activity of Tunisian *Nigella sativa* essential oil and thymoquinone. *Food Chemistry*, (2011), 129: 1469-1474.
37. Mohammed N.A., Effect of *Nigella Sativa* L. extracts against *Streptococcus mutans* and *Streptococcus mitis* in vitro. *J. Bagh. College. Dentistry*, (2012), 24(3): 154-157.
38. Emeka L.B., Emeka P.M., Khan T.M., Antimicrobial activity of *Nigella sativa* L. seed oil against multi-drug resistant *Staphylococcus aureus* isolated from diabetic wounds. *Pak. J. Pharm. Sci.*, (2015), 28(6): 1985-1990.
39. Rafati S., Niakan M., Naseri M., Anti-microbial effect of *Nigella sativa* seed extract against staphylococcal skin Infection. *Med. J. Islam. Repub. Iran.*, (2014), 8(28): 42.
40. Nawarathne N.W., Wijesekera K., Wijayarathne W.M.D.G.B., Napagoda M., Development of novel topical cosmeceutical formulations from *Nigella sativa* L. with antimicrobial activity against acne-causing microorganisms. *Sci. World J.*, (2019), 1-7.
41. Kocoglu E., Tayyar Kalcioğlu M., Uzun L., Zengin F., Celik S., Serifler S., Gulbay H., Gonullu N., In vitro investigation of the antibacterial activity of *Nigella sativa* Oil on some of the most commonly isolated bacteria in otitis media and externa. *Eurasian J. Med.*, (2019), 51(3): 247-51.

42. Mouhajir F., Pedersen J.A., Rejdali M., Towers G.H.N., Antimicrobial thymoquinones of Moroccan *Nigella Sativa* seeds detected by electron spin resonance. *Pharm. Biol.*, (1999), 37(5): 391-395.
43. Ferdous A.J., Islam S.N., Ahsan M., Hasan C.M., Ahmed Z.U., In Vitro Antibacterial Activity of the Volatile Oil of *Nigella sativa* Seeds against Multiple Drug-resistant Isolates of *Shigella* spp. and Isolates of *Vibrio cholerae* and *Escherichia coli*. *Phytotherapy Research*, (1992), 6: 137-140.
44. Piras A., Rosa A., Marongiu B., Porcedda S., Falconieri D., Dessì M.A., Ozcelik B., Koca U., Chemical composition and in vitro bioactivity of the volatile and fixed oils of *Nigella sativa* L. extracted by supercritical carbon dioxide. *Ind. Crop. Prod.*, (2013), 46: 317-323.
45. Randhawa M.A., Alenazy A.K., Alrowaili M.G., Basha J., An active principle of *Nigella sativa* L., thymoquinone, showing significant antimicrobial activity against anaerobic bacteria. *J. Intercult. Ethnopharmacol.*, (2017), 6(1): 97-101.
46. Chaieb K., Kouidhi B., Jrah H., Mahdouani K., Bakhrouf A., Antibacterial activity of Thymoquinone, an active principle of *Nigella sativa* and its potency to prevent bacterial biofilm formation. *BMC Complement. Altern. Med.*, (2011), 11(29): 1-6.
47. Hosseinzadeh H., Fazly Bazzaz B.S., Haghi M.M., Antibacterial Activity of Total Extracts and Essential oil of *Nigella Sativa* L. seeds in Mice. *Pharmacology online*, (2007), 2: 429-435.
48. Aljabre S.H.M, Randhawa M.A., Akhtar N., Alakloby O.M., Alqurashi A.M., Aldossary A., Antidermatophyte activity of ether extract of *Nigella sativa* and its active principle, thymoquinone. *J. Ethnopharmacol.*, (2005), 101: 116-119.
49. Sunita M., Meenakshi S., Chemical composition and antidermatophytic activity of *Nigella sativa* essential oil. *Afr. J. Pharm. Pharmacol.*, (2013), 7(20): 1286-1292.
50. Asdadi A., Harhar H., Gharby S., Bouzoubaâ Z., El Yadini A., Moutaj R., El Hadek M., Chebli B., Hassani L.M.I., Chemical composition and antifungal activity of *Nigella Sativa* L. oil seed cultivated in Morocco. *IJPSI.*, (2014), 3(11): 09-15.
51. Moghim H., Taghipoor S., Shahinfard N., Kheiri S., Panahi R., Antifungal effects of *Zataria multiflora* and *Nigella sativa* extracts against *Candida albicans*. *J. Herb. Med. Pharmacol.*, (2015), 4(4): 138-141.
52. Khan M.A.U., Ashfaq M.K., Zuberi H.S., Mahmood M.S., Gilani A.H., The in vivo antifungal activity of the aqueous extract from *Nigella sativa* seeds. *Phytother. Res.*, (2003), 17: 183-186.
53. Aftab, Yousaf A.Z., Javaid A., Riaz N., Younas A., Rashid M., Shamsheer H.B., Chahel A.A., Antifungal activity of methanolic extracts of *Nigella sativa* against *Fusarium oxysporum* and *Macrophomina phaseolina* and its phytochemical profiling by GCMS analysis. *Intl. J. Agric. Biol.*, (2019), 21: 569-576.
54. Sitara U., Niaz I., Naseem J., Sultana N., Antifungal effect of essential oils on in vitro growth of pathogenic fungi. *Pak. J. Bot.*, (2008), 40(1): 409-414.
55. Mahmoudvand H., Tavakoli R., Shariffar F., Minaie K., Ezatpour B., Jahanbakhsh S., Sharifi I., Leishmanicidal and cytotoxic activities of *Nigella sativa* and its active principle, thymoquinone. *Pharm. Biol.*, (2015), 53(7): 1052-1057.
56. Al-Turkmani M.O., Mokrani L., Soukkarieh C., Antileishmanial apoptotic activity of *Nigella sativa* L. essential oil and thymoquinone triggers on *Leishmania tropica*. *Indian J. Exp. Biol.*, (2020) 58: 699-705.
57. Mahmoudvand H., Shariffar F., Rahmat M.S., Tavakoli R., Dezaki E.S., Jahanbakhsh S., Sharifi I., Evaluation of antileishmanial activity and cytotoxicity of the extracts of *Berberis vulgaris* and *Nigella sativa* against *Leishmania tropica*. *J. Vector Borne Dis.*, (2014), 51: 294-299.

58. Jabbar E.A.K., AL-Aboody B.A., Jarullah B.A., Noori N., Isolation and molecular diagnosis of leishmania major and study activity of aqueous extract of plant *Nigella sativa* against the parasite in vitro. *IJPQA*, (2019), 10(1): 47-50.
59. Kadhim L.I., Al-Zubaidi M.T.S., AL Saegh H.A.H., Influence of dietary supplementation of *Nigella sativa* on experimental coccidiosis in broiler chickens. *JEZS*, (2018) 6(1): 652-656.
60. Sadek H.A., Abdel-Rahman S.M., Bakir H.Y., Arafa M.I., Ahmed A.A., Gareh A.A., Gaber M.M., The potential convention of garlic and black seed different extracts as an effective treatment of *Cryptosporidium* spp.: an experimental study. *J. Egypt. Soc. Parasitol.*, (2020), 50(3): 613-621.
61. Kabbashi A.S., Garbi M.I., Osman E.E., Dahab M.M., Koko W.S., Abuzeid N., Antigiardial and Cytotoxicity of Ethanolic Seed Extract of *Nigella sativa* (Linn) in Sudan. *JFPI*, (2015), 4(2): 66-72.
62. Kabbashi A.S., Osman E.E., Garbi M.I., Ahmed I.F., Saleh M.S., Badri A.M., Elshikh A.A., Abuzeid N., Koko W.S., Dahab M.M., In Vitro Antiprotozoal Activities and Cytotoxicity of Selected Sudanese Medicinal Plants. *Int. J. Biomed. Eng. Clin. Sci.*, (2017), 3(2): 6-13.
63. Sheikh B.Y., Taha M.M.E., Koko W.S., Abdelwahab S.I., Antimicrobial effects of thymoquinone on *Entamoeba histolytica* and *Giardia lamblia*. *Pharmacogn. J.*, (2016), 8(2): 168-170.
64. Ekanem J.T., Yusuf O.K., Some biochemical and haematological effects of black-seed (*Nigella sativa*) oil on *Trypanosoma brucei* infected rats. *Afr. J. Biotechnol.*, (2008), 7(2): 153-157.
65. Dyary H.O., Arifah A.K., Sharma R.S., Rasedee A., Mohd-Aspollah M.S., Zakaria Z.A., Zuraini A., Somchit M.N., Antitrypanosomal screening and cytotoxic effects of selected medicinal plants. *Trop. Biomed.*, (2014), 31(1): 89-96.
66. Mady R.F., El-Hadidy W., Elachy S., Effect of *Nigella sativa* oil on experimental toxoplasmosis. *Parasitol. Res.*, (2016), 115(1): 379-390.
67. Rayan H.Z., Wagih H.M., Atwa M.M., Efficacy of black seed oil from *Nigella sativa* against Murine Infection with Cysts of Me49 Strain of *Toxoplasma gondii*. *PUJ*, (2011), 4(2): 165-176.
68. Okeola V.O., Adaramoye O.A., Nneji C.M., Falade C.O., Farombi E.O., Ademowo O.G., Antimalarial and antioxidant activities of methanolic extract of *Nigella sativa* seeds (black cumin) in mice infected with *Plasmodium yoelli nigeriensis*. *Parasitol. Res.*, (2011), 108: 1507-1512.
69. Oyweri J., Mohammed A., Udu R., Gathirwa J., Too E., Omondi P., Kimani F., Hashim S., Abubakar L., In vitro and in vivo antimalarial activity of *Nigella sativa* L. Extracts. *J. Med. Plants Res.*, (2019), 13(19): 501-508.
70. Abdulelah H.A.A., Zainal-Abidin B.A.H., In Vivo Anti-malarial Tests of *Nigella sativa* (Black Seed) Different Extracts. *Am. J. Pharm. Toxicol.*, (2007), 2 (2): 46-50.
71. Udu R., Oyweri J., Gathirwa J., Antimalarial activity of *Nigella sativa* L. seed extracts and selection of resistance in *Plasmodium berghei* ANKA in a Mouse Model. *J. Pathog.*, (2021), 1-10.
72. Emeka P.M., Badger-Emeka L.I., Eneh C.M., Khan T.M., Dietary supplementation of chloroquine with *Nigella sativa* seed and oil extracts in the treatment of malaria induced in mice with *plasmodium berghei*. *Pharmacogn. Mag.*, (2014), 10(38): 357-362.
73. Sosiawan T.I., Linda W., Etty W., Anti-malaria study of *Nigella sativa* L. Seed Water Extract in *Mus musculus* mice Balb C strain in vivo. *Makara J. Sci.*, (2012), 16(3): 192-196.

74. Tonkal A.M.D., In vitro antitrichomonal effect of Nigella Sativa aqueous extract and wheat germ agglutinin. JKAU: Med. Sci., (2009), 16(2): 17-34.
75. Mahmoud M.A.E.A., Aminou H.A., Hashem H.A., Are the fatty acids responsible for the higher effect of oil and alcoholic extract of Nigella sativa over its aqueous extract on Trichomonas vaginalis trophozoites?. J. Parasit. Dis., (2016), 40(1): 22-31.
76. Al-Ammash M.S.J., Study the effect of alcoholic extract of Nigella sativa seeds on Trichomonas vaginalis in-vitro. Ibn Al-Haitham J. for Pure Appl. Sci., (2017), 03 (3): 10-18.
77. Aminou H.A., Alam-Eldin Y.H., Hashem H.A., Effect of Nigella sativa alcoholic extract and oil, as well as Phaseolus vulgaris (kidney bean) lectin on the ultrastructure of Trichomonas vaginalis trophozoites. J Parasit. Dis., (2016), 40(3):707-713.
78. Elkadery A.A.S., Elsherif E.A., Ezz Eldin H.M., Fahmy I.A.F., Mohammad O.S., Efficient therapeutic effect of Nigella sativa aqueous extract and chitosan nanoparticles against experimentally induced Acanthamoeba keratitis. Parasitol. Res., (2019), 118: 2443-2454.
79. Cheikh-Rouhou S., Besbes S., Hentati B., Blecker C., Deroanne C., Attia H., Nigella sativa L.: Chemical composition and physicochemical characteristics of lipid fraction. Food Chem., (2007), 101(2): 673-681.
80. Mohammed N.K., Abd Manap M.Y., Tan C.P., Muhiyaldin B.J., Alhelli A.M., Meor Hussin A.S., The effects of different extraction methods on antioxidant properties, chemical composition, and thermal behavior of Black Seed (Nigella sativa L.) oil. Evid. Based Complement. Alternat. Med., (2016), 2016: 6273817.
81. Piras A., Rosa A., Marongiu B., Porcedda S., Falconieri D., Dessì M.A., Ozcelik B., Koca U., Chemical composition and in vitro bioactivity of the volatile and fixed oils of Nigella sativa L. extracted by supercritical carbon dioxide. Ind. Crops. Prod., (2013), 46: 317-323.
82. Sudhir S.P., Padma Deshmukh, Verma H.N., Comparative study of antimicrobial effect of Nigella sativa seed extracts from different geographies. Int. J. Pharmacogn., (2016), 3(6): 257-264.
83. Kokoška L., Havlik J., Valterova I., Sovova H., Sajfrtova M., Marsik P., Antibacterial activity of Nigella sativa seed essential oil and effect of different extraction methods on content of its active principle, thymoquinone. Planta Med., (2006), 72(11): 083.
84. Kokoska L., Havlik J., Valterova I., Sovova H., Sajfrtova M., Jankovska I., Comparison of Chemical Composition and Antibacterial Activity of Nigella sativa Seed Essential Oils Obtained by Different Extraction Methods. J Food Prot., (2008), 71(12): 2475-2480.